MOLECULAR EPIDEMIOLOGY OF TUBERCULOSIS IN THE CZECH REPUBLIC, 2004: ANALYSIS OF M. TUBERCULOSIS COMPLEX ISOLATES ORIGINATING FROM THE CITY OF PRAGUE, SOUTH MORAVIA AND THE MORAVIAN-SILESIAN REGION

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SUMMARY

Objectives: To compare M. tuberculosis complex genotypes from representative regions of the Czech Republic in order to estimate changes in strain prevalence and in the extent of imported drug-resistant tuberculosis.

Methods: Primary M. tuberculosis complex isolates (n = 155) and follow-up isolates (n = 15) from 155 patients from the first half of 2004 (98 from Prague, 37 from South Moravia and 35 from the Moravian-Silesian region) were genotyped by IS6110-RFLP, spoligotyping, and partly by VNTR-genotyping.

Results: Based on IS6110-RFLP, 110 of 155 (71%) primary isolates were unique. Forty-five isolates (29 %) were found in 15 clusters comprising two to six patients and all but one cluster were also discriminated by MIRU-VNTR-genotyping. Four clusters comprised patients from different regions, and six were ongoing for several years. An indication of MDR-strain transmission was found in one instance. All four Beijing-type isolates with any resistance were associated with immigration from Eastern Europe.

Conclusions: The molecular epidemiological data of this period-prevalence, population based study and its comparison to earlier investigations point to a low extent of clustering between M. tuberculosis complex isolates in representative regions of the Czech Republic. Few clusters extending geographically and/or over several years were identified, providing a means for an in-depth analysis of risk factors of transmission. Beijing genotype isolates were shown to increase in prevalence to reach 6.5%. Drug resistant isolates of this genotype were associated with immigration of from Eastern Europe, although direct transmission of a resistant isolate was probable only in one of eleven cases.

Key words: tuberculosis, fingerprinting, epidemiology, Beijing-genotype

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INTRODUCTION

Molecular methods to study the epidemiology of tuberculosis have become common, since IS6110 restriction fragment length polymorphism (RFLP) for Mycobacterium tuberculosis was introduced in 1993 (32). Both the understanding of the natural history of tuberculosis (TB) and the clarification of transmission chains have been advanced through such molecular epidemiological studies (33). Based on genotyping studies, some strain-families, in particular the Beijing genotype, were postulated to be associated with higher pathogenicity (1, 8), and partly also with evolving drug resistance (8, 13). As a consequence, the introduction of multidrug-resistant (MDR) TB from countries of the former Soviet Union has become a constant threat for Central European countries like the Czech Republic (8, 12, 19, 21). E. g., the proportion of notified cases of TB in foreigners in the Czech Republic has increased from 5.9 % in 1997 to 14.1 % in 2004. Of the latter, patients from the neighbouring country Ukraine (n = 45) represented 30 % (3).

Molecular analyses of M. tuberculosis complex isolates, primarily IS6110-RFLP, have been employed in the Czech Republic on several instances, starting with a point-prevalence investigation of isolates from the City of Prague (14). Further studies focussed on a MDR-TB microepidemic (16), on TB in prisoners (16, 20), and on TB transmission in South Moravia, an area of particularly low TB incidence (17). The spoligotyping method (9) was employed...
to study the prevalence of strains families, especially the Beijing genotype, in Prague and South Moravia (18). A first comprehensive IS6110–RFLP analysis for the Czech M. tuberculosis population included 231 isolates or 73% and 90%, respectively, of all culture-positive TB cases from Prague and South Moravia, respectively, occurring in 1998 (ref. 21). The results suggested a proportion of cases attributable to recent transmission similar to other Central European countries with low TB incidence rates, although a few clusters were shown to be prevalent in both study regions. Only sporadic importation of TB from foreigners was noted at that time.

The present study surveys the Czech M. tuberculosis population from the first half of 2004 from three representative regions: the City of Prague, South Moravia, and the Moravian-Silesian region. The study areas are geographically, demographically and epidemiologically different areas: the metropolitan area of Prague, the national capital, counts 1.2 million inhabitants and with 12.0 per 100,000 had the second highest TB incidence rate in the country in 2004 (ref. 2). For comparison, the incidence rates in 2004 were 10.3 for South Moravia and 9.6 for the Moravian-Silesian region (2). From 1998 until 2004, Czech notification data have shown a stable number of MDR cases which has remained between 2 and 10 cases per year (0.5 and 2 %, respectively, of all tested patients’ isolates) (7). Several resistant or MDR isolates were included in this study, and for the first time genotyping data and drug sensitivity testing results were correlated on a national scale.

This analysis is still based on IS6110-RFLP. However, a forthcoming standard method for TB molecular epidemiology (22, 30, 31), mycobacterial interspersed repeat elements (MIRU) genotyping or – in more general terms – variable number tandem repeat (VNTR) genotyping, was applied for the first time to a representative sample of Czech isolates. This should allow connecting previous with contemporary data.

MATERIAL AND METHODS

Patients’ Demographic Characteristics

Data were collected by the culturing laboratories and checked against the notification reports of TB cases in the Czech Republic. Depersonalized initials were used outside the respective culturing laboratory to identify patients.

M. tuberculosis Complex Isolates

Isolates obtained by standard culturing techniques in one of the participating laboratory between January 1st and June 30th, 2004, were included in the study. For 15 patients, follow-up isolates were included as blinded internal controls throughout the fingerprinting procedure. The identity as M. tuberculosis complex was confirmed by biochemical and/or genotypic tests. For genotyping, all isolates were subcultured on Löwenstein-Jensen medium. Drug-sensitivity testing for first line drugs was done by the proportion method on solid media according to standard techniques in the participating laboratories.

Genotyping Procedures

DNA fingerprinting was performed on subcultured isolates at the Innsbruck Medical University. Genomic DNA was isolated as described (32) and used for all fingerprinting methods. IS6110–RFLP was performed according to Van Embden et al. (32) using the ECL Direct Nucleic Acid Labelling and Detection System (Amersham Biosciences, Chalfont, UK). The patterns were analyzed with Gelcompar® version 4.2 (Applied Math, Sint-Martens-Latem, Belgium) by the unweighted pair group method using arithmetic averages and the Dice coefficient for similarity with 1% band position tolerance. A cluster was defined as a group of isolates having an identical IS6110–RFLP, cluster size was calculated by the N method, i.e. including the index case. Banding patterns from clusters found in the 1998 study (21) were compared manually to the patterns of cluster strains in this study, as different software had been employed in the earlier study. Spoligotyping was carried out according to Kamerbeek et al. (9) using first generation membranes (Isogen Bioscience BV, Maarsen, the Netherlands). The octal codes for spoligotypes were determined as proposed (5). Names for global spoligotypes are according to the international spoligotyping database SpolDB4 (4). MIRU-VNTR genotyping was performed on all RFLP-clustered isolates by 12 single PCRs as described (29), based on the method of Philip Supply (22, 30). MIRU copy numbers corresponding to the respective band size were taken from published copy number tables (www.lif.fr/mirus/mirus.html). Six additional VNTR loci (VNTR numbers: 0424, 3690, 0577, 4156, 2401, 1982) were tested as described (31).

RESULTS

Epidemiological Characteristics of Patients

One hundred and ninety-two isolates from Prague, South Moravia, and the Moravian-Silesian region (n = 103, 43, and 45, respectively) were submitted for genotyping. After correcting for isolates lost during recultivation or analysis and for the follow-up isolates, 170 isolates from 155 individuals were genotyped. The 155 patients recruited in 6 months represent 60.5 % of all culture positive patients reported in these areas in the whole year 2004, and 23.3 % of all Czech TB cases notified in that period, respectively (3). Basic demographic data of these individuals are displayed in Fig. 1 and Table 1. The mean age of all study patients was 51 years (range: 19–85 years), female patients tended to be older than males. There was no gross difference in age distribution between the three study areas, although the male:female ratio, being 2.37 for all patients, was 3.63 in South Moravia. Twenty-nine patients (19 %) were born outside Czech or Slovak territory, and most foreign-born study patients were situated either

<table>
<thead>
<tr>
<th>Geographic origin of isolates</th>
<th>All patients, No.</th>
<th>Mean age (range)</th>
<th>Male: female ratio</th>
</tr>
</thead>
<tbody>
<tr>
<td>Prague</td>
<td>83</td>
<td>53 (19-85)</td>
<td>1.96</td>
</tr>
<tr>
<td>South Moravia</td>
<td>37</td>
<td>47 (18-82)</td>
<td>3.63</td>
</tr>
<tr>
<td>Moravian-Silesian region</td>
<td>35</td>
<td>51 (24-88)</td>
<td>2.5</td>
</tr>
<tr>
<td>All</td>
<td>155</td>
<td>51 (18-88)</td>
<td>2.37</td>
</tr>
</tbody>
</table>

Table 1. Characteristics of 155 patients with genotyped M. tuberculosis complex isolates, according to region of origin, Czech Republic 2004
in Prague (n = 14) or in a district in the Moravian-Silesian region (n = 9; all asylum seekers). Foreign-born patients were significantly younger than native Czechs or Slovaks (Fig. 1). Homeless patients (n = 3) or patients in jail (n = 4) represented only small fractions. The demographic data of nine patients (all native Czechs and including one case of MDR-TB) whose isolates were lost during analysis were not significantly different from those of the patients with genotyped isolates.

Cluster Analysis by IS6110 –RFLP

All 170 isolates in the analysis were typeable by RFLP. Three isolates contained less than 6 copies of IS6110, but showed unique patterns. The 15 follow-up isolates that served as internal standards contained 6 copies of IS6110.

Cluster / No. | IS6110-RFLP pattern | MIRU code (canonical 12 + new 6)
---|---|---
C 1 3 | | 22331531321
C 2 4 | | 22332531325
C 3 2 | | 22332531324
C 4 2 | | 223323143321 223323
C 5 2 | | 223323143321 223323
C 6 3 | | 223325173533
C 7 2 | | 22332513633
C 8 2 | | 22332517423
C 9 2 | | 22725113223
C 10 2 | | 22332513323
C 11 4 | | 22332513323 234332
C 12 4 | | 22332513324 234336
C 13 4 | | 22332513324 234336
C 14 3 | | 22332513324 234336
C 15 2 | | 22332513324

Fig. 4. IS6110-RFLP patterns and MIRU-VNTR codes for prototype isolates representing the 15 clusters (C1 through C15) found among 155 M. tuberculosis complex isolates from the Czech Republic, 2004. The number of isolates (n) per cluster is indicated. The MIRU code lists the copy numbers at the 12 canonical loci 2-4-10-16-20-23-24-26-27-31-39-40. For some isolates, six additional VNTR loci (sequentially: VNTR numbers 0424, 0577, 2401, 3690, 4156, and 1982) are given. Codes for different RFLP-clusters that were identical or nearly identical are underlined. The arrow points to the one band distinguishing C4 and C5.
controls for the reculturing and fingerprinting process yielded identical patterns for 14 pairs, and one pattern that differed by one additional faint band from that of the corresponding primary isolate. The IS6110 copy number among the 155 patients’ isolates showed a distribution skewed to the right, with high-copy number isolates being of the Beijing genotype and the main peak centered around ten IS6110 copies (Fig. 2). This distribution was similar in all three regions (mean copy numbers range: 10.0 to 10.4), but different for clustered isolates (range 8 – 22 copies, mean 12.1) and non-clustered ones (range 1 – 19 copies, mean 9.5).

Analysed together, the 155 patients’ isolates were grouped into 110 individual types and 45 isolates found in 15 clusters, and non-clustered ones (range 1 – 19 copies, mean 9.5). The numbers for cluster isolates are given by the N method (i.e., including the index case); n.a., not applicable.

Table 2. Clustering by IS6110-RFLP of 155 patients’ M. tuberculosis isolates, according to region of origin, Czech Republic 2004

<table>
<thead>
<tr>
<th>Geographic origin of isolates</th>
<th>All isolates collected for fingerprinting including follow-ups (n)</th>
<th>All typed isolates excluding follow-ups (n)</th>
<th>Isolates in RFLP-based cluster n (% of all)</th>
<th>RFLP clusters (only isolates with this origin) (n)</th>
<th>Minimal – maximal number of isolates per cluster</th>
</tr>
</thead>
<tbody>
<tr>
<td>Prague</td>
<td>103</td>
<td>83</td>
<td>25 (30.1 %)</td>
<td>5</td>
<td>2-6</td>
</tr>
<tr>
<td>South Moravia</td>
<td>43</td>
<td>37</td>
<td>11 (29.7 %)</td>
<td>3</td>
<td>2-3</td>
</tr>
<tr>
<td>Moravian-Silesian region</td>
<td>35</td>
<td>35</td>
<td>9 (25.7 %)</td>
<td>3</td>
<td>2</td>
</tr>
<tr>
<td>Clusters with isolates from &gt;1 region</td>
<td>n.a.</td>
<td>n.a.</td>
<td>n.a.</td>
<td>4</td>
<td>2-6</td>
</tr>
<tr>
<td>All</td>
<td>181</td>
<td>155</td>
<td>45 (29 %)</td>
<td>15</td>
<td>2-6</td>
</tr>
</tbody>
</table>

Table 3. Most frequent (No. of isolates >2) and world-wide distributed shared types (ST) by spoligotyping among 155 patients’ isolates from the Czech Republic, 2004

<table>
<thead>
<tr>
<th>ST name</th>
<th>Octal code</th>
<th>No. of isolates</th>
</tr>
</thead>
<tbody>
<tr>
<td>Haarlem3</td>
<td>777777777720771</td>
<td>29</td>
</tr>
<tr>
<td>T1</td>
<td>777777777760771</td>
<td>21</td>
</tr>
<tr>
<td>Beijing</td>
<td>000000000037771</td>
<td>11</td>
</tr>
<tr>
<td>CZ-ST 1</td>
<td>677777777720771</td>
<td>9</td>
</tr>
<tr>
<td>Haarlem1</td>
<td>7777777774020771</td>
<td>7</td>
</tr>
<tr>
<td>CZ-ST 2</td>
<td>774037777760700</td>
<td>5</td>
</tr>
<tr>
<td>CZ-ST 3</td>
<td>77777760000000</td>
<td>5</td>
</tr>
<tr>
<td>CZ-ST 4</td>
<td>77777743760771</td>
<td>4</td>
</tr>
<tr>
<td>T2</td>
<td>777777777720731</td>
<td>4</td>
</tr>
<tr>
<td>T3</td>
<td>777773777760771</td>
<td>4</td>
</tr>
<tr>
<td>CZ-ST 5</td>
<td>77777757777777</td>
<td>3</td>
</tr>
<tr>
<td>CZ-ST 6</td>
<td>777777807760771</td>
<td>3</td>
</tr>
<tr>
<td>Haarlem2</td>
<td>000000000420771</td>
<td>3</td>
</tr>
<tr>
<td>LAM7</td>
<td>777777404760771</td>
<td>1</td>
</tr>
<tr>
<td>Vietnam type</td>
<td>77777774413771</td>
<td>1</td>
</tr>
</tbody>
</table>

MIRU-VNTR Genotyping of Cluster Prototype Isolates

For all 45 RFLP-clustered isolates the copy numbers for the twelve canonical MIRU-type VNTR loci were assessed. For 29 of these 45 isolates, i.e., for clusters with more than 3 isolates and for any clusters with an identical 12-loci profile, the copy numbers in six additional VNTR loci were determined. Fifteen different MIRU-VNTR combinations were found and their distribution was nearly congruent with RFLP clustering (see Fig. 4). Clusters C4 and C5 which differed by only one band were identical by all 18 VNTR loci. C11 and C13, clearly distinguishable by RFLP, were only separated by 2 out of 6 additional VNTR loci. Within cluster C12, 2 of 6 isolates were minimally different by one copy in 1 of 18 loci.

Correlation of Genotyping and Drug Sensitivity data

Of 170 isolates, 158 had been found fully susceptible against first line drugs. For one isolate the DST result could not be procured, and 11 showed a resistant phenotype: 3 were resistant against INH, 3 against INH and SM, and 5 were MDR with the resistance patterns INH+RMP (n = 2), INH+RMP+SM (n = 2), (Table 2). The three clusters that comprised more than 3 isolates were two clusters in Prague with 4 and 6 isolates, respectively, and one occurring in all three regions, also with 6 isolates. The latter and further three clusters comprising patients at more than one study region contributed one third (15 of 45) of all patients with clustered isolates. No statistically significant differences between patients with clustered isolates and patients with non-clustered isolates were found regarding sex, age, homelessness, or nationality. Czech or Slovak nationals were included in 14 of the 15 clusters, three of these included also foreign-born patients.

Analysis by Spoligotyping

Spoligotyping of all 170 isolates revealed 68 different spoligotypes, or shared types (STs) according to (4). Forty-five of the 68 STs were seen only once. All isolates clustered by IS6110–RFLP also shared the same ST. On the other hand, however, the STs associated with 14 of the 15 RFLP clusters were also found in non-clustered isolates. Eighty-one isolates (48 % of all isolates) showed a world-wide encountered prototype ST, predominantly of the Haarlem, T or Beijing family (Table 3). Only 6 other STs, termed CZ-ST 1 through 6, were present in at least three isolates each, or in altogether 29 isolates (see Table 3).
or INH+ RMP+SM+EMB (n = 1). By IS6110-RFLP, only the two INH+RMP+SM resistant isolates were clustered: They were isolated from an elder Czech and a young homeless Georgian patient, respectively, both from the Moravian-Silesian region, but not from the same district. Contact tracing information revealed that both had stayed in the same hospital at the same time. One INH-monoresistant isolate was identical by genotyping to a susceptible one from the same region, all other resistant isolates were unclustered. Seven resistant isolates were obtained from patients from the former Soviet Union and 4 from Czech citizens. The seven immigrants were younger than Czech patients with resistant TB (mean age 27 vs. 51 years; p < 0.01). Furthermore, four patients with different resistant isolates lived in one home for asylum seekers in the Moravian-Silesian region. The location of this home is the reason that this region has a significantly higher proportion of resistant isolates than the other two regions (14% vs. 4%; p < 0.05). Of altogether 11 Beijing-type isolates, 4 were associated with drug resistance, and all were isolated from immigrants from Eastern Europe (Table 4). In contrast, non-resistant Beijing-type isolates were only found in native Czech (n = 5) or Vietnamese (n = 2) individuals.

### DISCUSSION

In terms of TB incidence, the Czech Republic can be regarded as in between the Western European countries and Eastern European former Soviet Union member states (7). As a Central European country, the country is exposed to migration along the economical West-East divide and flight from warfare in former Soviet Union member states. These trends are considered to contribute in particular to the observed rise in drug resistant TB (13). The present study therefore encompassed for the first time the Moravian-Silesian region on the Czech-Ukrainian border with a state-operated home for asylum seekers, together with the urban area of Prague and the rural South Moravian region. Although representing only a circumspect period of time, the study allowed making qualitative and quantitative estimates of strain prevalence and respective changes to the situation several years before.

The IS6110-RFLP clustering analysis revealed 29% of isolates in clusters altogether and without significant differences between the three study regions. This is in keeping with the data presented by Kurepina et al. (21) which showed 32% clustering for isolates from 1998 (28% and 35% in Prague and South Moravia, respectively, compared to 30% for each of the two regions in 2004). Furthermore, native Czech patients have a relatively high mean age (54 yrs). The limited duration of the study does not allow to estimate the proportion of TB cases attributable to recent transmission, however; it suggests that recent transmission of TB is not a large problem in the Czech Republic. As might have been expected, some cluster patterns were found both in 1998 and 2004: Five out of nine Prague clusters in the 1998 analysis were seen again in 2004, comprising 58% (in 1998) and 64% (2004) of the Prague patients with clustered isolates, respectively, which is remarkable both in the extent and steadiness. In 2004, however, these clusters also included 7 isolates from outside Prague, suggesting an expansion from the capital to the other regions. Alternatively, keeping in mind the short and discontinuous observation period, these strains could have been circulating for a long time throughout the country: this hypothesis would be supported by the old age of native Czech TB patients, which is independent of cluster status (mean age 52 vs. 55 years in patients with clustered or unclustered isolates). On the contrary, there was little continuity between the cluster prototypes in South Moravia: of the 15 cluster prototypes in 1998, only the largest one, including patients from South Moravia and Prague at both sampling times, was present in 2004. All clusters in 2004 with isolates from South Moravia only, or from the Moravian-Silesian region only, were small (on average 2.2 patients including index case).

For the first time, the drug resistance status of prevalent isolates could be compared to the genotype on a national scale. Of 11 resistant isolates (5 MDR and 6 with any other resistance), only two were identical. As both had stayed in the same hospital at the same time, a transmission in either direction appears possible, whereas a laboratory contamination could be excluded. This low proportion of clustered resistant isolates may most likely be explained as a result of multiple individual treatment failure, or of multiple imports from abroad or other Czech regions, or of both. In this regard, the distribution of resistant and susceptible isolates of the Beijing genotype and the change of Beijing-type prevalence over time was assessed. All four resistant isolates found with immigrant Eastern Europeans had the Beijing-genotype, although clearly different by RFLP, and no further resistant Beijing-type isolate was identified. Such evidence for multiple, parallel import of MDR-Beijing type strains has been documented also in other Central European countries (13). The prevalence of the Beijing-type among Czech isolates appears to have increased.

### Table 4. Occurrence of Beijing-genotype isolates, Czech Republic 2004

<table>
<thead>
<tr>
<th>Geographic origin of isolates</th>
<th>All typed strains (one per patient)</th>
<th>All Beijing-genotype isolates</th>
<th>Beijing-genotype isolates in clusters</th>
<th>Foreign origin in patients with Beijing-genotype isolates</th>
<th>Beijing-genotype isolates with any drug resistance (or MDR)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Prague</td>
<td>83</td>
<td>4</td>
<td>0</td>
<td>3</td>
<td>1 (1)</td>
</tr>
<tr>
<td>South Moravia</td>
<td>37</td>
<td>5</td>
<td>5 (2 clusters)</td>
<td>2</td>
<td>1 (0)</td>
</tr>
<tr>
<td>Moravian-Silesian region</td>
<td>35</td>
<td>2</td>
<td>0</td>
<td>2</td>
<td>2 (0)</td>
</tr>
<tr>
<td>All</td>
<td>155</td>
<td>11</td>
<td>5</td>
<td>6</td>
<td>4 (1)</td>
</tr>
</tbody>
</table>

- 7.1% of all isolates
- 45% of all Beijing-genotype isolates
- 55% of all Beijing-genotype isolates
- 35% of all Beijing-genotype isolates
from 3% in 1998 (21) to 6.5% in this study. Whether the increased number of – susceptible – Beijing-type isolates in native Czech patients is merely the result of more than a decade of probably increased contact with such strains or reflects their postulated increased pathogenicity, cannot be decided in this place. Altogether, however, evidence can be demonstrated for the role of importation of resistant TB from Eastern Europe, which is often, but not always associated with the Beijing genotype. Other factors associated with MDR-TB like homelessness (23) or imprisonment appear to be of less importance in this study sample. Of the seven cases of TB in prisoners or homeless, all but one (a prisoner of Moldovan nationality with a double resistant isolate) had individual genotypes there were susceptible to first-line drugs. However, that these factors are relevant, has been documented earlier in the Czech Republic in an outbreak of MDR-TB comprising homeless, imprisoned, or immigrant patients in the Czech Republic (16).

The distribution of strain families in the Mycobacterium tuberculosis population is apparently stable over the last years. Unsurprisingly, isolates of the Haarlem and T-family, respectively, were seen most often (Table 3). Other world-wide prevalent STs were rare, with the exception of the Beijing type. Notably, a large number of individual STs (n = 45) was recorded, as was the case in 1998 (ref. 21). This adds to the evidence of a generally very heterogeneous Mycobacterium tuberculosis population, interestingly without a single isolate of Mycobacterium bovis or Mycobacterium caprae among the 401 isolates in both studies. This is in contrast to the experience of Germany or Austria where rates around 0.5% of “bovine” isolates in humans have been observed (11, 28). Although Mycobacterium caprae is a frequent cause of animal TB in Middle Europe (6, 24, 25, 29), bovine tuberculosis in cattle has apparently been decimated in the country very efficiently (26).

Finally, we compared the clustering results by the current TB fingerprinting standard technique, IS6110-RFLP, with those of the forthcoming standard, MIRU-VNTR genotyping (30, 31). All clusters formed by RFLP but one were distinguishable by MIRU-VNTR. Two large clusters separated by 2 “additional” out of 18 VNTR loci, supporting the need of more than the 12 canonical loci for population-based VNTR typing. On the other hand, the microheterogeneity in cluster C12 was only reflected by VNTR typing. We have not applied the MIRU-VNTR method to the RFLP-unclustered isolates, however, and the discrimination between those isolates has to be determined by future work. In studies conducted elsewhere, MIRU-VNTR was shown to discriminate sufficiently well between epidemiologically unrelated isolates (22, 31). We conclude that the VNTR data from this study are promising in connecting data from earlier and future studies on TB epidemiology from the Czech Republic and other European countries.

Acknowledgements
This study was inspired by the meetings of the European Union Concerted Action “New generation markers and techniques for the epidemiology and control of tuberculosis” (QLK2-CT-2000-00630). We thank Dr. Milan Šlosárek from the National Reference Laboratory of Mycobacterial Diseases for technical assistance and fruitful consultations and Prof. Ivo Pavlík, Veterinary Research Institute Brno, for many helpful discussions.

REFERENCES
The editors are affiliated with the Harvard Medical School and with the Boston University School of Medicine, Boston, Massachusetts. The list of contributors comprises 32 international experts in multiple areas of immunology— they come from USA, Canada, Europe, and Lebanon. As indicated in the preface, there are many reasons why microbes have outwitted humans’ ability to control infectious diseases. But at the center of this struggle between microbes and the humans is the immune system. Without an immune system, there is only a short period of survival, terminating by overwhelming infection. The volume is arranged into six sections, it embraces 27 chapters. Each chapter is composed of several subchapters, and concludes with a summary and a list of suggesting readings. All the chapters have standardized thematic and structural aspects to provide critical information in a comprehensive style.

Introducing Section I “Function and Composition of the Immune System” (chapters 1 through 5) provides an overview of immunity, furthermore examined are innate immunity, cells of the immune system, organs and tissues of the immune system, and the complement. Section II “Antibodies” (chapters 6 through 10) is devoted to molecular genetics of antibody diversity, to antigens, antigenicity, and immunogenicity, to antibody-antigen interactions and measurements of immunologic reactions, to B-lymphocyte activation, and antibody production. Section III “Cellular Immunity” (chapters 11 through 15) explores the major histocompatibility complexity, antigen processing and presentation, the T-cell receptor, T-cell maturation and activation, and cellular communication. Section IV “Immunologic Effector Systems and Immunity to Infection” (chapters 16 through 21) focus on mediated immunity, mucosal immunity, immunity to bacterial infections, immunity to viruses, immunity to parasitic and fungal infections, and to vaccines and vaccination. Section V “Immune System Dysfunctions: Deficiencies” (chapters 22 through 24) is concerned with immunology and AIDS, clinical and genetic perspectives in primary immunodeficiency disorders, and cancer and the immune system. Concluding section VI “Immune System Dysfunctions: Overactivity” (chapters 25 through 27) provides a look at hypersensitivity, autoimmunity and disease, and transplantation and immunity. In addition, there are 7 appendices presented in form of tabular reviews or figures. Discussed are CD antigens, antigen names with CD designation equivalents, cytokines, chemokines and their receptors, cell types and immune related functions, historical timeline of immunology, time course of a typical immune response, and comparison of relative sizes of cells, structures, and molecules that are relevant to infection and immunity. The volume is comprehensively illustrated by remarkable and attractive, mostly colour figures depicting basic molecular and cellular components of immune systems, cells and tissues, and schematic immunological processes.

Immunology, Infection, and Immunity complements traditional views and dogmas about immunology with today’s cutting edge ideas and data from experiments describing how the immune system works, some of which are challenging and changing long-held beliefs about the function of the immune system.

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