Supplementary Table S1. Strains of *Acinetobacter berezniae* sp. nov. and *Acinetobacter guillouiae* sp. nov.

Culture collections: ATCC, American Type Culture Collection, Manassas, VA, USA; CCM, Czech Collection of Microorganisms, Brno, Czech Republic; CCUG, Culture Collection, University of Göteborg, Sweden; LMG, Bacteria Collection, Laboratorium voor Microbiologie Gent, Gent, Belgium; CIP, Collection de l’Institut Pasteur, Institut Pasteur, Paris, France. ANC and NIPH are strain designations used by A. Nemec; LUH and RUH are strain designations used by L. Dijkshoorn. CZ, Czech Republic; NL, the Netherlands; in, inpatient; out, outpatient.

<table>
<thead>
<tr>
<th>Strain Specimen Donor Reference Accession number</th>
<th>Location and year of isolation</th>
<th>DNA–DNA reassociation rpoB 16S rRNA gene</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>Acinetobacter berezniae</em> sp. nov. (n=16)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>LMG 1003T (=NIPH 521T =LUH 13182T =ATCC 17924T =CIP 70.12T =69T*)</td>
<td>Wound (human) Before 1960</td>
<td>1</td>
</tr>
<tr>
<td>CCGU 28628 (=RUH 2223 =NIPH 2260 =113:2*)</td>
<td>Wound (human) Malmo, Sweden, 1980</td>
<td>I. Tjernberg</td>
</tr>
<tr>
<td>RUH 2222 (=NIPH 2274 =198*)</td>
<td>Urine (human) Malmo, Sweden, 1980</td>
<td>I. Tjernberg</td>
</tr>
<tr>
<td>NIPH 870 (=LUH 10180)</td>
<td>Urine (human, in) Medical Unit, Budéjovice, CZ, 1997</td>
<td>M. Horníková</td>
</tr>
<tr>
<td>NIPH 1741 (=LUH 10183)</td>
<td>Sputum (human, in) Plzeň, CZ, 2001</td>
<td>T. Bergerová</td>
</tr>
<tr>
<td>LUH 2634 (=NIPH 2524)</td>
<td>Sink (hospital environment) Enschede, NL, 1995</td>
<td></td>
</tr>
<tr>
<td>LUH 6973 (=NIPH 2532)</td>
<td>Faeces (human, out) Leiden, NL, 2000</td>
<td></td>
</tr>
<tr>
<td>LUH 9667 (=NIPH 2542 =86307)</td>
<td>Eye (rabbit) Dublin, Ireland</td>
<td></td>
</tr>
<tr>
<td>NIPH 3 (=LUH 10179)</td>
<td>Hospital environment Prague, 1991</td>
<td>J. Vránková</td>
</tr>
<tr>
<td>NIPH 1050 (=LUH 10181)</td>
<td>Urine (human, in) Sedlčany, CZ, 1998</td>
<td>P. Jelík</td>
</tr>
<tr>
<td>NIPH 1054 (=LUH 10182)</td>
<td>Faeces (human, out) Pribram, CZ, 1998</td>
<td>P. Jelík</td>
</tr>
<tr>
<td>LMG 5624 (=NIPH 2532 =R1-82T)</td>
<td>Sewage Denmark, 1997</td>
<td>L. Guardabassi</td>
</tr>
<tr>
<td>LMG 5852 (=NIPH 2531)</td>
<td>Sputum (human, in) Leiden, NL, 1999</td>
<td></td>
</tr>
<tr>
<td>LMG 7438 (=NIPH 2535 =118F1C)</td>
<td>Blood (human) Coimbra, Portugal, 1998</td>
<td>G. da Silva</td>
</tr>
<tr>
<td>LMG 7832 (=NIPH 2537 =V0112893F)</td>
<td>Wound (human) Utrecht, NL, 2001</td>
<td>J. Wagenaar</td>
</tr>
<tr>
<td>LMG 8524 (=NIPH 2539 =130:30:21)</td>
<td>Clinical specimen (human) Heerlen, NL, 2003</td>
<td>Wagenvoort</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th><em>Acinetobacter guillouiae</em> sp. nov. (n=17)</th>
<th></th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td>LMG 988T (=NIPH 522T =LUH 13183T =ATCC 11717T =CIP 63.46T =73T*)</td>
<td>Sewage Before 1951</td>
<td>1</td>
</tr>
<tr>
<td>RUH 2234 (=NIPH 2272 =174*)</td>
<td>Contact lens Malmo, Sweden, 1980</td>
<td>I. Tjernberg</td>
</tr>
<tr>
<td>RUH 2236 (=NIPH 2273 =51T)</td>
<td>Urine (human) Malmo, Sweden, 1980</td>
<td>I. Tjernberg</td>
</tr>
<tr>
<td>NIPH 682 (=LUH 13118)</td>
<td>Blood (human, in) Medical Unit, Budéjovice, CZ, 1997</td>
<td>O. Hausner</td>
</tr>
<tr>
<td>RUH 2860 (=NIPH 2127 =58B*)</td>
<td>Wound (human) Malmo, Sweden, 1980</td>
<td>I. Tjernberg</td>
</tr>
<tr>
<td>LMG 267 (=NIPH 2169 =225T)</td>
<td>Sputum (human) Odense, Denmark, 1988-9</td>
<td>P. Gerner-Smidt</td>
</tr>
<tr>
<td>RUH 1050 (=NIPH 82O =LMD 81.109)</td>
<td>Not known</td>
<td>J. van der Toorn</td>
</tr>
<tr>
<td>A23 (=NIPH 2689 =LUH 10615 =CIP 107475)</td>
<td>Activated sludge Albury, Australia</td>
<td>E. Carr</td>
</tr>
<tr>
<td>LMG 5606 (=NIPH 2525 =PAUG7)</td>
<td>Freshwater with sediment Jutland, Denmark, 1997</td>
<td>L. Guardabassi</td>
</tr>
</tbody>
</table>


<table>
<thead>
<tr>
<th>Strain</th>
<th>Specimen</th>
<th>Location and year of isolation</th>
<th>Donor</th>
<th>Reference</th>
<th>Accession number</th>
<th>DNA–DNA reassociation rpoB 16S rRNA gene</th>
</tr>
</thead>
<tbody>
<tr>
<td>LUH 5653 (=NIPH 2529)</td>
<td>Blood (human, in)</td>
<td>Leiden, NL, 1999</td>
<td></td>
<td></td>
<td>FJ754453</td>
<td></td>
</tr>
<tr>
<td>LUH 7830 (=NIPH 2536 =V0112205†)</td>
<td>Eye (cat)</td>
<td>Utrecht, NL, 2001</td>
<td>J. Wagenaar</td>
<td></td>
<td>FJ754457</td>
<td></td>
</tr>
<tr>
<td>LUH 4560 (=NIPH 769 =A130d†)</td>
<td>Soil (barley field)</td>
<td>Porthtowan, Cornwall, UK, 1993–4</td>
<td>H. Seifert</td>
<td></td>
<td>FJ754432</td>
<td></td>
</tr>
<tr>
<td>LUH 6980 (=NIPH 2680)</td>
<td>Faeces (human, out)</td>
<td>Leiden, NL, 2000</td>
<td></td>
<td></td>
<td>FJ754461</td>
<td></td>
</tr>
<tr>
<td>NIPH 991 (=LUH 7854)</td>
<td>Ear swab (human, out)</td>
<td>Sedlčany, CZ, 1998</td>
<td>P. Ježek</td>
<td>6</td>
<td>FJ754435</td>
<td></td>
</tr>
<tr>
<td>ANC 3626 (=LUH 13178)</td>
<td>Soil (beech forest)</td>
<td>CZ, 2007</td>
<td>J. Wagenaar</td>
<td></td>
<td>FJ754429</td>
<td></td>
</tr>
<tr>
<td>LUH 7013 (=NIPH 2681)</td>
<td>Faeces (human, out)</td>
<td>Leiden, NL, 2000</td>
<td>L. Krňová</td>
<td></td>
<td>FJ754462</td>
<td>FM177775</td>
</tr>
<tr>
<td>CCM 4725 (=NIPH 2408 =LUH 10204)</td>
<td>Raw milk</td>
<td>North Moravia, CZ</td>
<td></td>
<td></td>
<td>FJ754449</td>
<td></td>
</tr>
</tbody>
</table>

* Strain designation used in the reference publication.
† Strain designation used by the donor.

References


Supplementary Fig. S1. Dendrogram of cluster analysis of AFLP fingerprints of 16 strains of *Acinetobacter bereziniae* sp. nov., 17 strains of *Acinetobacter guillouiae* sp. nov., and 30 strains representing all known (genomic) species of the genus *Acinetobacter*. AFLP was performed as described by Nemec et al. (2001), with the following steps: simultaneous digestion of DNA with two restriction endonucleases (EcoRI and MseI) and adapter ligation, PCR with a Cy5-labelled EcoRI+A primer and an MseI+C primer (A and C are selective nucleotides), separation of amplified fragments with the ALF Express system (Amersham Biosciences) and cluster analysis of fingerprints with the BioNumerics software release 4.6 (Applied Maths) using Pearson’s product moment correlation coefficient (r) for similarity calculation and UPGMA for clustering. The arrow indicates the species delineation level.

Reference


Supplementary Fig. S2. MALDI-TOF MS analysis of strains of *Acinetobacter bereziniae* sp. nov. and *Acinetobacter guillouiae* sp. nov. (a) Segment of MALDI-TOF mass spectra of 11 strains each of *A. bereziniae* sp. nov. and *A. guillouiae* sp. nov., with highlighted peaks characteristic of a particular species. (b) Score-orientated dendrogram derived from MALDI-TOF mass spectra of the these 22 strains and four strains representing other *Acinetobacter* species.